A RARE OCCURRENCE OF MATCHED OTOLITHS AND ASSOCIATED SKELETAL REMAINS OF *APOGON TOWNSENDI* (OSTEICHTHYES) FROM THE CALOOSAHATCHEE FORMATION (LOWER PLEISTOCENE) OF FLORIDA

Gary L. Stringer¹, Richard C. Hulbert Jr.², Dirk Nolf³, Paul Roth⁴, and Roger W. Portell⁴

ABSTRACT

A matched pair of otoliths (right and left saccular otoliths) and associated skeletal remains (n = 107) of Apogon townsendi (belted cardinalfish) were obtained in unconsolidated sediment from inside the valves of an articulated scallop Carolinapecten eboreus. The scallop specimen was collected in Hendry County, Florida, from the lower Pleistocene Caloosahatchee Formation, approximately 1.7 to 2.1 Ma. The recovery of this vertebrate material is highly significant because no skeletal remains of bony fish with in situ or associated otoliths are known from the Gulf or Atlantic coasts of the United States. Furthermore, the specimen represents the first fossil record of the family Apogonidae and the genus *Apogon* from Florida and the first report of the species Apogon townsendi in the fossil record. The length of the fossil Apogon townsendi was determined to be 4.7 cm based on the linear relationship between fish length and otolith length and utilizing modern specimens of the species for comparison and analysis. The length of the fossil Apogon townsendi indicated that it was an adult fish upon death (> 2.1 cm). Although several taphonomic scenarios are considered, including commensalism, it is believed that the apogonid died in close proximity to the empty scallop shell, which was followed by fairly rapid washing in of sediment with the fish into the valves of the scallop (i.e., sediment trapping). This determination is based on several factors including the biology, distribution, and behavior of extant *Apogon townsendi*. The presence and preservation of the two matched saccular otoliths and the large number of associated, fragile skeletal remains in the scallop shell suggest that the apogonid was not ingested and excreted by a piscivorous predator.

Key words: *Apogon townsendi*; Caloosahatchee Formation; Florida; otolith; Pleistocene; taphonomy.

Published On-line: May 8, 2017

Open Access Download at https://www.flmnh.ufl.edu/bulletin/publications/

ISSN 2373-9991

Copyright © 2017 by the Florida Museum of Natural History, University of Florida. All rights reserved. Text, images and other media are for nonprofit, educational, or personal use of students, scholars, and the public. Any commercial use or republication by printed or electronic media is strictly prohibited without written permission of the museum.

Museum of Natural History, Hanna Hall, University of Louisiana at Monroe, Monroe, Louisiana 71209 USA < stringer@ulm.edu

²Division of Vertebrate Paleontology, Florida Museum of Natural History, University of Florida, Gainesville, Florida 32611-7800 USA

³Royal Belgian Institute of Natural Sciences, 29, rue Vautier, 1000 Brussels, Belgium

⁴Division of Invertebrate Paleontology, Florida Museum of Natural History, University of Florida, Gainesville, Florida 32611-7800 USA

INTRODUCTION

Osteological fish remains with otoliths *in situ* or with associated otoliths are rarely found. Nolf (1985) listed 23 fossil taxa of otoliths associated with skeletal material in which the otoliths were adequately preserved to be taxonomically beneficial. Since Nolf (1985), other skeletal remains with *in situ* or associated otoliths have been reported in North America, Europe, and Asia (Fierstine et al., 2012; Nolf, 2013:16–18; Schwarzhans et al., 2016:2–3).

There are at least three previous cases where fish skeletal remains with associated otoliths were found inside mollusk shells. Schwarzhans and Weiler (1971) described otoliths of a Pristigenys species (under the name of "Trigla arambourgi?"; see comments in Taverne and Nolf, 2010:233) from sediment inside a shell of the gastropod Natica crassatina collected in the Reupelian "Meeressand" of the Mainz Basin, Germany. Stinton (1980:209) mentioned otoliths of Pristigenys dentifer (a synonym of Pristigenys rutoti) collected with associated bones in a shell of the gastropod Clavilithes macrospira from the English Bartonian, and Taverne and Nolf (2010) reported Pristigenys skeletal material and associated otoliths recovered from the inside of several nautilid shells from the Sands of Lede (middle Eocene) of Belgium at the locality of Meldert, Belgium. Two Pristigenys species (P. rutoti and P. hermani) were recognized from different nautilid shells.

Studies of fish skeletons with otoliths (*in situ* or associated) in North America are even scarcer but have been reported from the western United States by Huddleston and Takeuchi (2002, 2006), Takeuchi and Huddleston (2008), and Fierstine et al. (2012). The importance of skeletal remains with associated otoliths for the purpose of taxonomic identifications and relationships was emphasized by Takeuchi and Huddleston (2008). No reports of skeletal remains with *in situ* or associated otoliths are known from the Gulf or Atlantic coasts of the United States, which increases the significance of the matched otoliths associated with osteological material from the Caloosahatchee Formation.

LOCALITY DATA AND STRATIGRAPHY

The otoliths and associated skeletal remains were collected at UF locality HN017 located in Clewiston in Hendry County, Florida (26.75389, -80.93389 WGS84)(Fig. 1). The collected material is from the lower Pleistocene Caloosahatchee Formation, approximately 1.7 to 2.1 Ma (Kolbe et al., 2011 and references therein). The deposit is predominately marine in origin with minor interbedded freshwater marls. The areal extent of the Caloosahatchee Formation (with a few exceptions) is southern peninsular Florida. Lithologies consist of shelly quartz sands, calcarenites, and limestones. Much of the deposit is unconsolidated except areas converted to limestones or calcarenites by subaerial weathering or action of groundwater (DuBar, 1974). Caloosahatchee Formation index mollusks found at UF locality HN017 include Siphocypraea problematica (Heilprin, 1886), Diodora carolinensis (Conrad, 1875), and Cerithioclava caloosaense (Dall, 1892). DuBar (1974) interpreted the paleoecology of the Caloosahatchee Formation to reflect deposition in a shallow, marginal sea and adjacent low-lying coastal areas.

MATERIAL AND METHODS

The matched pair of otoliths (right and left saccular otoliths) and associated skeletal material were obtained from unconsolidated sediment found inside an articulated set of valves of the scallop Carolinapecten eboreus (Conrad, 1833)(UF/IP 256774; Fig. 2). The scallop was 8.6 cm in width and 8.9 cm in length. It was collected by the late George and Wylda Stephens and was part of a collection transferred to the Florida Museum of Natural History from the Virginia Tech Department of Geosciences. The otoliths and associated bones were discovered during the water screening of the unconsolidated sediments in the scallop shell. The skeletal remains were disarticulated prior to processing but may have been fragmented further during the screening. The sediment was water screened using U.S. sieve sizes #14 (1.41 mm openings), #35 (0.50 mm openings), and #70 (0.20 mm openings). The otoliths and associated skeletal remains were recovered in

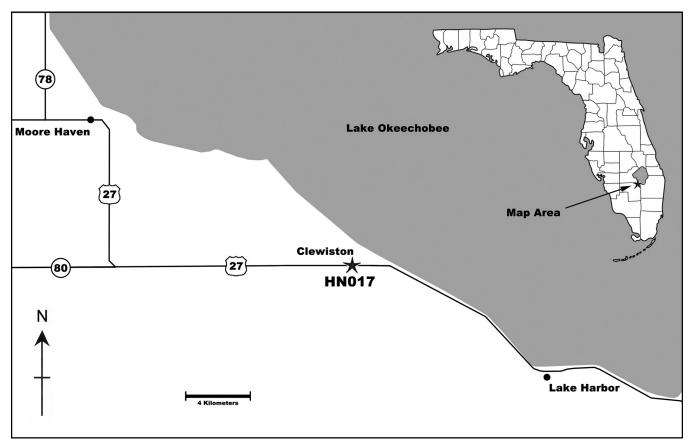


Figure 1. Map showing the location of UF locality HN017 in Hendry County, Florida, which produced the matched pair of fish otoliths and associated skeletal remains

the largest residue (sieve size #14). No other vertebrate remains were found within the scallop.

The vertebrate remains (UF 319000) consisted of well-preserved right and left saccular otoliths and associated skeletal material consisting of 107 disarticulated bones. Although much of the bony material was broken and fragmented, various skeletal elements were recognized including fin spines, vertebrae, premaxilla, maxilla, dentary, quadrate, posttemporal, and numerous fragmented bones of the skull. Institutional abbreviations are as follows: IRSNB, Institut royal des Sciences naturelles, Brussels, Belgium; UF, Division of Vertebrate Paleontology, FLMNH, Gainesville, Florida, USA; UF/IP, Division of Invertebrate Paleontology, FLMNH, Gainesville, Florida, USA; FLMNH, Florida Museum of Natural History, Gainesville, Florida, USA; YPM, Yale Peabody Museum of Natural History, New Haven, Connecticut, USA.

SYSTEMATIC PALEONTOLOGY

Key morphological features, such as the sulcus consisting of a wide ostium and narrower cauda with a collicular crest near the crista inferior, the prominent depression above the sulcus, the shape of the otolith, and the prominent anterodorsal dome, of the matched pair of saccular otoliths from the Caloosahatchee Formation clearly indicate an apogonid. Böhlke and Chaplin (1968) and McEachran and Fechhelm (2005) indicated the following Recent apogonids as occurring in southern Florida and the northern Caribbean: Apogon (12 species), Astrapogon (2 species), Cheilodipterus (1 species), Paroncheilus (1 species), and Phaeoptyx (2 species). Apogonid otoliths are often distinguished by subtle differences in the sulcus and the outline of the saccular otoliths and require well-preserved and abundant specimens (Nolf, 2013). Fortunately, the Caloosahatchee Formation otoliths were almost

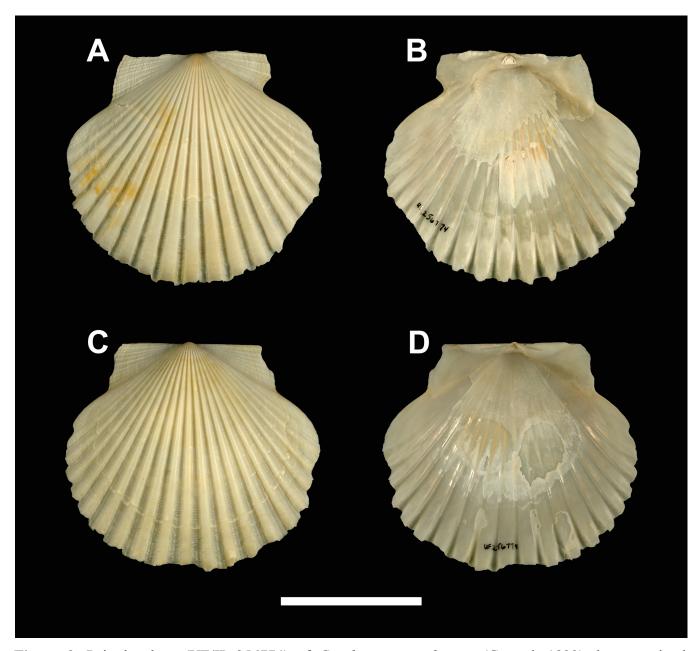


Figure 2. Paired valves (UF/IP 256774) of *Carolinapecten eboreus* (Conrad, 1833) that contained sediments with matched pair of fish otoliths and associated skeletal material. A. Exterior view of right valve. B. Interior view of right valve. C. Exterior view of left valve. D. Interior view of left valve. Scale Bar equals 5 cm.

perfectly preserved and allowed for identification. The Caloosahatchee Formation otoliths were compared with the otoliths of Recent apogonids that occur in southern Florida and the Caribbean from the IRSNB collection. They agree best with those of *Apogon townsendi*, which was illustrated by Nolf and Bajpai (1992)(Fig. 3). Stringer (1992) reported a similar apogonid from a late Pleistocene

mudlump island in the Gulf of Mexico. Higher level taxonomy follows Nelson et al. (2016).

Class OSTEICHTHYES
Subclass ACTINOPTERYGII
Order KURTIFORMES
Family APOGONIDAE Günther, 1859
Subfamily APOGONINAE Günther, 1859
Genus *APOGON* Lacepède 1801

APOGON TOWNSENDI (Breder, 1927)

Figures 4–5

Type specimens.—YPM 460, holotype; YPM 463 to 465, paratypes, taken from Saddle Rock, Washerwoman Cut, Bahamas (Eschmeyer et al., 2017).

Saccular otolith description.—Both of the otoliths (from the left and right side; UF 319000) measure 3.5 mm in length and 2.85 mm in height (Fig. 4). The length to height ratio is 1.23. The outline of the otolith is primarily oval with a conspicuous anterodorsal dome. The margins tend be sinuate with the ventral margin more equally broad. The inner face is only very slightly convex and is characterized by a clearly delimited sulcus that is located approximately in the center. The distinct ostium is approximately 25% wider and 20% longer than the cauda. The ostium is slightly slanted toward the dorsal margin, whereas the cauda is essentially horizontal. The crista superior is well developed over part of the ostium and cauda. Colliculum is present in the ostium and cauda. The dorsal area is characterized by a pronounced oval depression located above the junction of the ostium and the cauda. The ventral area is characterized by a shallow ventral groove that is parallel to the ventral margin. The outer face is markedly more convex than the inner face and primarily smooth with some slight undulations.

Skeletal elements description.—Although no skeletal remains of A. townsendi were available, lateral and medial views of the maxilla, premaxilla, dentary, angular, and quadrate of the Recent apogonid Apogon menesemus in Dye and Longenecker (2004) were compared to the bones found in association with the otoliths. The Caloosahatchee skeletal remains, especially the diagnostic premaxilla, compared very favorably with the Recent apogonid, but the actual species determination was based on the otoliths. However since the matched otoliths were found in the scallop valve, then it is highly probable that the skull of the fish was present. Since the only vertebrate remains in the scallop shell were the otoliths and the apogonid bones, it appears to be a valid assumption that the bones were A. townsendi. As noted previously,

the majority of the skeletal remains of *A. townsendi* (UF 319000) were broken and highly fragmented. However, some skeletal elements were recognizable and are briefly described.

One of the best-preserved skull elements is the right premaxilla; possible fragments of the left premaxilla are also present. This is fortunate since

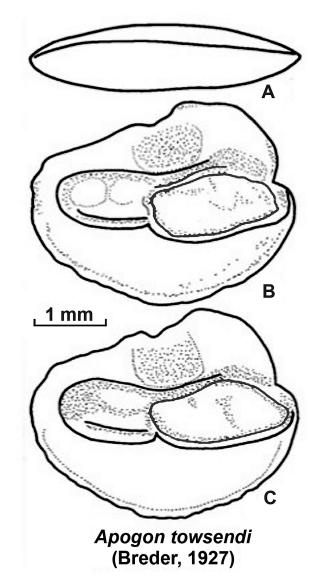


Figure 3. Left saccular otoliths from two Recent specimens of *Apogon townsendi* (Breder, 1927) from off Mona Island, Lesser Antilles (Coll. IRSNB). Dorsal view of first specimen (A). Inner face of first specimen (B). Inner face of second specimen (C). Figure modified from Nolf and Bajpai (1992).

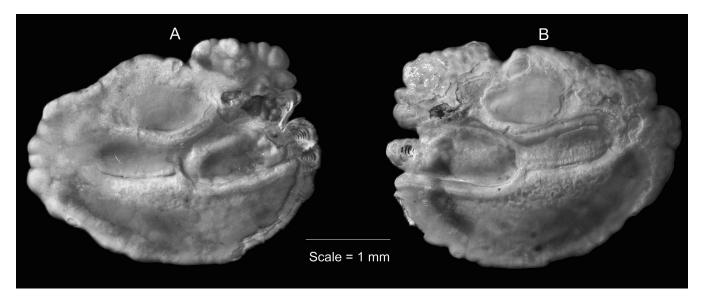


Figure 4. Matched pair of fossil otoliths (UF 319000) from *Apogon townsendi* from the Caloosahatchee Formation (lower Pleistocene), Hendry County, Florida (UF locality HN017). Left saccular otolith (A) and right saccular otolith (B).

premaxillae have been utilized in identification of marine fishes (Watt, 1997), and Dye and Longenecker (2004) refer to it as the second most distinctive bone in the fish skull. The right premaxilla measures 4.72 mm in length and is 2.19 mm in anterior width and 1.75 mm in posterior width (Fig. 5F). In lateral view, the anterior is characterized by the ascending process and the articular process. Numerous highly distinctive features of the Caloosahatchee premaxilla match Recent apogonid premaxillae (Dye and Longenecker, 2004). These include: 1) although incomplete, the ascending process is quite thin and delicate; 2) the interprocess notch between the ascending process and articular process is a very diagnostic, acute v-shape; 3) the shape of the articular process; 4) the outline and shape between the articular process and the postmaxillary; 5) the postmaxillary is quite expanded in length and height, and the height is about the same as the height of the articular process; and 6) the caudal process is broken but appears that it would be quite short. All of these features on the Caloosahatchee premaxilla match those of apogonids (Cannon, 1987; Watt, 1997; Dye and Longenecker, 2004). In medial view, the anterior has a distinct articular surface and obvious fossa, and these features are almost identical in the aforementioned Recent apogonid.

In ventral view, the anterior has a pronounced occlusal surface with buccal and lingual socket rows. The socket rows on the anterior tend to be smaller in diameter than at the posterior. The ramus is slightly curved and increases in width towards the posterior as found in the apogonid of Dye and Longenecker (2004). There are fragments of what appear to be parts of the occlusal surface of the left premaxilla. A fragment of an occlusal surface measuring 1.3 mm by 1.4 mm had approximately 120 tooth bases (Fig. 5G). The fragment probably came from the premaxilla or the dentary.

A fairly significant portion of one of the maxilla is preserved and measures 7.49 mm in length (Fig. 5I). The anterior end is broken. The maxilla is rounded at the anterior and becomes broader and flatter towards the posterior. The posterior has some breakage also. A part of the quadrate, measuring 3.18 mm in length, was recovered. Although incomplete, several features were discernible. The anterior end is 1.1 mm in width and is characterized by two condyles (lateral and mesial). The preopercular process runs along the ventral of the quadrate and is fairly complete. The process has a concavity in the middle (preopercular groove). The dorsal (ectopterygoid margin) is broken irregularly but is much thinner than the preopercular process.. Two

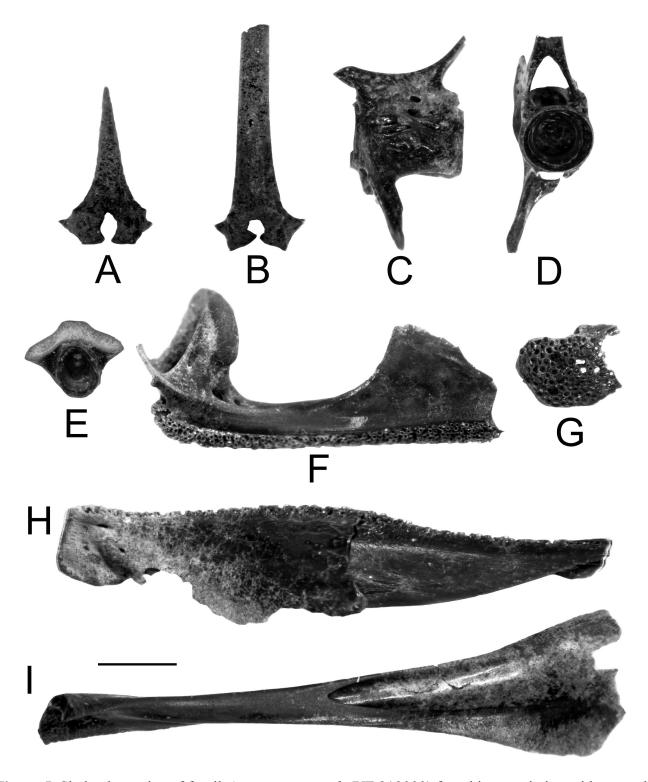


Figure 5. Skeletal remains of fossil *Apogon townsendi* (UF 319000) found in association with a matched pair of saccular otoliths in the Caloosahatchee Formation (lower Pleistocene), Hendry County, Florida (UF locality HN017). Anterior views of two dorsal spines (A, B). Lateral view of caudal vertebra (C). Anterior view of caudal vertebra showing partial neural and hemal arches (D). Anterior view of first anterior abdominal (atlas) vertebra showing articular surfaces for attachment to the basioccipital (E). Lingual lateral view of right premaxilla (F). Oral view of an occlusal surface fragment (G). Lingual lateral view of right dentary (H). Lingual lateral view of right maxilla (I).

incomplete, narrow bones with lengths of 4.58 mm and 4.21 mm and widths of approximately 0.60 mm are believed to be portions of the posttemporals. The proximal ends of the bones are broken, while the distal ends appear to be mainly complete. The bones are more rounded towards the proximal and become wider towards the distal. The distal end is flattened with a concavity (spatula-like).

There are two adjacent portions of the right dentary with a total length of 7.10 mm (Fig. 5H). Unfortunately, the preservation of the dentary is extremely fragile, and part is missing. The dentary is slightly curved, and the internal and external walls are clearly visible. The most distinct feature is the occlusal surface with socket rows for teeth. The socket rows tend to narrower and smaller in diameter at the anterior, near the symphysis. A deep, oval-shaped foramen (mental foramen) is visible on the labial lateral.

Nine spines (primarily dorsal but some possibly anal) were identified and ranged in length from 4.14 to 1.40 mm. Although the extant *A. townsendi* possesses seven dorsal and two anal spines (Snyder and Burgess, 2016), three of the recovered spines are medial sections, and it is not known if they represent a single spine or more. The only complete spine measured 1.90 mm in length and 0.60 mm at the proximal articulation (Fig. 5A). The distal end is slightly curved and quite aciculate. Another spine, which was 2.90 mm in length and 0.62 mm at the proximal articulation, is complete except for the distal end (Fig. 5B). The longest spine, which is 4.14 mm in length, is missing the proximal end (i.e., the articulation).

Fourteen vertebrae were recognized, most consisting of only the centra. All of the centra were distinctly amphicoelous. The best-preserved is a caudal vertebra measuring 1.43 mm in length and 2.80 mm in height, which included the arches (Fig. 5C). Both the neural and haemal spines are preserved with the enclosed arches (Fig. 5D). The neural and haemal spines are missing on three other caudal vertebra (1.35–1.43 mm in length), but the bases of the spines are visible. Two others (1.00–1.10 mm in length) appear to be abdominal vertebrae based on the presence of the remains of neural spines and parapophyses. One of these is the

first anterior abdominal vertebra or atlas. It is 1.10 mm in length and 1.00 mm in height, and the neural spine is broken. In addition to being quite compressed, the vertebra has distinct articular surfaces for attachment to the basioccipital at the base of the skull (Fig. 5E).

RESULTS

As noted in the Systematic Paleontology section, the salient morphological features of the Caloosahatchee Formation saccular otoliths agreed best with those of A. townsendi, and Nolf and Bajpai (1992) illustrated left otoliths from two Recent A. townsendi collected off Mona Island in the Lesser Antilles. The Recent fish were approximately 5.1 cm and 5.3 cm in body length with corresponding saccular otoliths of 3.7 mm and 3.8 mm in length (Coll. IRSNB). An approximate length of the fossil A. townsendi from the Caloosahatchee Formation can be determined because the relationship between the length of the fish and the length of the otolith is linear (Harvey et al., 2000; Mendoza, 2006; Rypel, 2008; Jawad et al., 2011). The matched pair of otoliths from the Caloosahatchee Formation are both 3.5 mm in length and would represent a fish of approximately 4.7 cm in length. Of course, the number of Recent specimens available limits this approximation, but it does provide a viable length for the fossil A. townsendi from the Caloosahatchee Formation. A determination of the length of the living fish provides important information with regards to the feasibility of certain taphonomic processes. Studies by Baldwin et al. (2011) using molecular data (DNA barcoding) determined that Recent A. townsendi 1.1 cm and smaller were larvae, while juveniles were 1.2 to 2.1 cm in length. This would indicate that the Caloosahatchee Formation fossil A. townsendi at 4.7 cm in length would be an adult, but still much smaller than the valves of Carolinapecten eboreus in which it was found.

The Caloosahatchee skeletal remains, especially the highly diagnostic premaxilla, compared quite well with the Recent apogonid *A. menesemus* and indicate an apogonid. Although the skeletal material indicates an apogonid, no comparative material was available for *A. townsendi*. Therefore, the actual species determination was based on the

otoliths of Recent *A. townsendi*. However, the skull of the fish was most likely present in the valves of the scallop given the matched pair of otoliths. The only vertebrate remains in the scallop shell were the otoliths and the apogonid bones, and it appears quite likely that the bones were also of *A. townsendi* and the same individual.

DISCUSSION

BIOLOGY, HABITAT, AND BEHAVIOR OF APOGON TOWNSENDI

Most apogonids are small, carnivorous reef fishes that commonly live in close association with reef invertebrates and are found primarily in tropical and subtropical areas (Nelson et al., 2016). Apogon townsendi follows these generalizations (Robins and Ray, 1986). Lieske and Myers (1994) and Gilmore and Fraser (2015) characterized the species as marine and reef-associated with a depth range of 3-55 m. Böhlke and Chaplin (1968) reported that A. townsendi was found in waters from 2.4–27.4 m with few shallower than 6.1 m. They also noted that the species was rather abundant on patch reefs in the 12.2-18.3 m depth range in the Nassau area of the Bahamas. Dennis et al. (2005) also reported A. townsendi from the reef biotype. Harborne et al. (2012) in a study of cryptobenthic fish in reef environments in the U.S. Virgin Islands found that A. townsendi was the most characteristic species in sheltered reef areas with <50% dead coral cover. A. townsendi is often found near drop-offs and may be found solitary or in groups in caves or holes (Lieske and Myers, 1994; Snyder and Burgess, 2106). It is often seen hovering in the spines of long-spined sea urchins (Gilmore and Fraser, 2015). Dominici-Arosemena and Wolf (2005) classified A. townsendi as a predator that hunts macrofauna and a mobile invertebrate feeder, whereas Halpern and Floeter (2008) and Gilmore and Fraser (2015) classified it as a midwater nocturnal planktivore. Based on the various references, it is probable that the species feeds on invertebrates as well as zooplankton.

Böhlke and Chaplin (1993), Claro (1994), and Randall (1996) listed *A. townsendi* in the Caribbean, Gulf of Mexico, and southeast U.S. Continental Shelf, respectively. The species is indicated as present in the Atlantic, Western-Central major

fishing area (#31) according to the Food and Agriculture Organization (FAO) of the United Nations major fishing areas (2017) with an approximate range of 5°–35° N and 40°–90° W. The reported distributions of Recent *A. townsendi* would include the fossil locality in Hendry County, Florida, during the early Pleistocene.

Most Recent apogonids are less than 10 cm in total length, and this would certainly increase the chances of being preserved in a bivalve (Nelson et al., 2016). Rivaton and Bourret (1999) illustrated otoliths from 15 different Recent *Apogon* species representing juveniles and adults. They ranged from 3.7 cm to 9.2 cm in total length with otoliths that ranged from 2.9 mm to 6.75 mm in length. The maximum known size of *A. townsendi* is about 6.5 cm (McEachran and Fechhelm, 2005; Snyder and Burgess, 2016).

FOSSIL RECORD OF APOGON

The oldest specimens referred to the genus Apogon in the scientific literature are from the middle Eocene of Pakistan, India, and England and are represented by otoliths (Gingerich et al., 1979; Kemp, 1985; Nolf and Bajpai, 1992). Gingerich et al. (1979) noted otoliths of Apogon sp. from the Domanda Formation in Pakistan while Nolf and Baipai (1992) reported otoliths of Apogon sp. 4 from the Harudi Formation in India. Kemp (1985) listed Apogon boulei and Apogon bellovacinus from the Bracklesham Group in England. A middle Eocene age for Apogon seems very reasonable given that comprehensive molecular studies have shown a date of approximately 45 Ma for the Apogonidae (Betancur-R. et al., 2013). Other morphology- and molecular-based systematic revisions of apogonids (e.g., Bergman, 2004; Mabuchi et al., 2014) have indicated that A. townsendi is a valid species in the family Apogonidae.

The specimen UF 319000 represents the first fossil record of the family Apogonidae and the genus *Apogon* from Florida. Neither were listed in the two largest fossil bony fish assemblages from the early Pleistocene of Florida, Richardson Road Shell Pit 6A (34 species; Emslie et al., 1996) and Leisey Shell Pit 1A (50 species; Scudder et al., 1995). Otoliths within the size range of UF 319000

were recovered at Richardson Road Shell Pit 6A (Emslie et al., 1996), so that collecting bias was not a factor in their absence. Instead this is likely related to unsuitable habitat for apogonids at these two sites. This is the first fossil record anywhere of the species *A. townsendi*.

TAPHONOMIC CONSIDERATIONS

The occurrence of otoliths and associated skeletal remains believed to belong to the same species and individual of A. townsendi in the articulated shells of Carolinapecten eboreus may be the result of a commensal relationship between the fish and the scallop. However, commensal relationships that are not manifested by abnormalities or distinct structures in the preserved remains of the host are difficult to establish in the fossil record (Zullo and Chivers, 1969). The best-known examples of commensalism of fish with mollusks, specifically bivalves, occur in several species of the Carapidae (Parmentier and Das, 2004). Smith (1955) noted Carapus margaritiferae in South African waters as being "taken from inside clams at Durban." Mahadevan (1961) reported the same pearlfish species as being collected in the bivalve Pteria from the East Indies and the Gulf of Mannar in India. Trott (1981) described the hosts of several species of carapids as bivalves. Nazar et al. (2011) indicated that the hosts of the pearlfish Onuxodon margaritiferas were limited to bivalves (Pteria penguin and Pycnodonta hyotis). Most adults of Carapus use their host for shelter and leave it to hunt, but other pearlfishes are free-living (Echiodon) and parasitic (Encheliohis) according to Trott (1970), Parmentier et al. (2000), and Parmentier and Das (2004), In addition to the carapids, some of the gobiids display a type of commensal relationship with burrowing invertebrates. For example, the Eastern Pacific Lepidogobius lepidus often occupies the burrows of invertebrates on intertidal mud flats (Hieb, 2000).

McEachran and Fechhelm (2005) observed that a number of Gulf of Mexico apogonids live within the body cavities of other organisms such as sponges and mollusks without causing harm to them, while others are closely associated with sea urchins. One of the apogonids, *Astrapogon stel*-

latus or conchfish, has a commensal relationship with the queen conch shell Lobatus gigas and with the bivalve Atrina rigida (Cervigón, 1993). The relationship of this species with L. gigas was noted by Plate (1908), who reported that Astrapogon stellatus emerged from the shell only at night to feed on small crustaceans. Randall (1967) specified that A. stellatus was a commensal in the mantle cavity of the queen conch. McEachran and Fechhelm (2005) also reported that another species of Astrapogon (A. puncticulatus) is often found in empty shells and that Phaeoptyx conklini is associated with empty conch shells. So, there are several apogonids commensal or associated with bivalves in the Gulf of Mexico.

As noted earlier, A. townsendi is often reported hovering in long-spined sea urchins and is in intimate contact with invertebrates, especially in reef settings. The positioning of the fish within the sharp spines of the sea urchin could provide some protection and perhaps even camouflage from predators. If commensalism is defined as a relationship between two kinds of organisms in which one obtains food or other benefits from the other without damaging or benefiting it, then A. townsendi could be considered commensal with the sea urchin on a limited basis (i.e., the amount of time spent in the sea urchin's spines). However, Albins and Lyons (2012), Snyder and Burgess (2016), and other references herein identify A. townsendi as a nocturnal, hole-dwelling species found primarily in reefs. Furthermore, there are no known reports of commensalism of A. townsendi with bivalves, but its proximity with bivalve species in the reef setting is well established.

Another taphonomic scenario for consideration is *A. townsendi* actually occupying the empty scallop shell with subsequent death and rapid sediment infilling. As noted above, several Gulf of Mexico and Caribbean apogonids consistently occupy empty bivalve shells. While *A. townsendi* has not been reported as occupying empty shells, it is a possibility since the species is hole dwelling and may have sought temporary shelter in the shell. Another alternative is sediment trapping or washing in of the fish into the empty scallop shell. This is a feasible consideration because complete,

delicate carapaces of crabs similar in size to A. townsendi have been described within articulated bivalves (Zullo and Chivers, 1969; Gašparič et al. 2015). This type of taphonomic process would most likely require death of the fish in close proximity to the empty scallop shell followed by fairly rapid washing in of sediment with the fish into the valves of the scallop (i.e., sediment trapping). Subsequent burial of the scallop assisted in preserving the unique assemblage. The otoliths do not appear to have been exposed to any type of invertebrate settlement on the sea bottom as described by Stringer (2016) and were washed into the scallop intact in the apogonid's skull. It is possible that the fossil A. townsendi was the prev of a piscivorous predator (ingested and excreted) but this is considered unlikely based on several factors. There is no indication of the corrosive action of digestion on the otoliths. Furthermore, it is highly unlikely that the delicate skeletal bones were preserved in a coprolite and subsequently extracted from the calcium fluoride phosphate matrix

CONCLUSIONS

The excellent preservation of the matched pair of otoliths from the Caloosahatchee Formation allowed them to be attributed to an Apogon townsendi of about 4.7 cm in length and was an adult fish at death. The skeletal remains, especially the premaxilla, compared extremely well to an apogonid and are believed to be from the same individual as the otoliths. Based on the biology, distribution, and behavior of living A. townsendi, the taphonomic process that led to this rare and unusual assemblage is believed to have involved the death of the apogonid near or within the empty scallop shell followed by fairly rapid washing in of sediment into the scallop valves. The preservation of the saccular otoliths and the large number of associated, fragile skeletal remains believed to belong to the same individual fish in the scallop shell suggest that the apogonid was not ingested and excreted by a piscivorous predator.

ACKNOWLEDGMENTS

Appreciation is extended to the late George and Wylda Stephens who collected the specimen of Carolinapecten eboreus that contained the studied material. S. Llyn Sharp of the Virginia Tech Department of Geosciences transferred the Stephens Collection to the FLMNH, and Sean Roberts (FLMNH) assisted with Figures 1 and 2. The late Eugenia Böhlke allowed Dirk Nolf to dissect some apogonid otoliths from fishes in the collection of the Academy of Natural Sciences, Philadelphia, and to incorporate them in the otolith reference collection of the Institut Royal des Sciences Naturelles de Belgique in Brussels, Belgium. We especially thank D. Bell (University of Louisiana at Monroe Museum of Natural History) for his work on the preparation of the plates of the otoliths and skeletal material. This manuscript was greatly improved by reviewers Dr. B. Welton (New Mexico Museum of Natural History and Science, Albuquerque) and Dr. W. Schwarzhans (Natural History Museum of Denmark, Copenhagen) and volume co-editor Jason Bourque (FLMNH). This is University of Florida Contribution to Paleobiology 833.

LITERATURE CITED

Albins, M., and P. Lyons. 2012. Invasive red lion-fish *Pterois volitans* blow directed jets of water at prey fish. Marine Ecology Progress Series 448:1–5.

Baldwin, C., B. Brito, D. G. Smith, E. Weight, and E. Escobar-Briones. 2011. Identification of early life-history stages of Caribbean *Apogon* (Perciformes: Apogonidae) through DNA barcoding. Zootaxa 3133:1–36.

Betancur-R. R., R. E. Broughton, E. O. Wiley, K. Carpenter, J. López, C. Li, N. Holcroft, D. Arcila, M. Sanciangco, J. Cureton II, F. Zhang, T. Buser, M. Campbell, J. Ballesteros, A. Roa-Varon, S. Willis, W. Borden, T. Rowley, P. C. Reneau, D. Hough, G. Lu, T. Grande, G. Arratia, and G. Ortí. 2013. The tree of life and a new classification of bony fishes. PLOS Currents Tree of Life. 2013 April 18 (last modified: 2013 April 23). Edition1.doi:10.1371/currents.tol.53ba26640df0ccaee75bb165c8c26288.

Bergman, L. M. R. 2004. The cephalic lateralis system of cardinalfishes (Perciformes: Apogonidae) and its application to the taxonomy and

- systematics of the family. Ph.D. Dissertation. University of Hawaii, Honolulu. 373 p.
- Böhlke, J., and C. Chaplin. 1968. Fishes of the Bahamas and Adjacent Tropical Waters. The Academy of Natural Sciences of Philadelphia, Livingston Publishing Company, Wynnewood, Pennsylvania, 771 p.
- Böhlke, J., and C. Chaplin. 1993. Fishes of the Bahamas and Adjacent Tropical Waters. Second edition. University of Texas Press, Austin, 857 p.
- Breder, C. M., Jr. 1927. Scientific results of the first oceanographic expedition of the "Pawnee" 1925. Fishes. Bulletin of the Bingham Oceanographic Collection Yale University 1(1):1–90.
- Cannon, D. 1987. Marine fish osteology: a manual for archaeologists. Simon Fraser University Publication 18:1–133.
- Cervigón, F., 1993. Los Peces Marinos de Venezuela. Volume 2. Fundación Científica Los Roques, Caracas, Venezuela, 497 p.
- Claro, R., 1994. Características generales de la ictiofauna. Pp. 55–70 *in* R. Claro, ed. Ecología de los peces marinos de Cuba. Instituto de Oceanología Academia de Ciencias de Cuba and Centro de Investigaciones de Quintana Roo.
- Conrad, T. A. 1833. On some new fossil and Recent shells of the United States. American Journal of Science and Arts 23:339–346.
- Conrad, T. A. 1875. Descriptions of new genera and species of fossil shells of North Carolina, in the state cabinet at Raleigh. Appendix A. Pp. 1–28 *in* W. C. Kerr, ed. Report of the Geological Survey of North Carolina.
- Dall, W. H. 1892. Contributions to the Tertiary fauna of Florida. Part II. Sreptodont and other gastropods, concluded. Transactions of the Wagner Free Institute of Science of Philadelphia 3:201–473.
- Dennis, G., W. Smith-Vaniz, P. Colin, D. Hensley, and M. McGehee. 2005. Shore fishes from islands of the Mona Passage, Greater Antilles with comments on their zoogeography. Caribbean Journal of Science 41(4):716–743.

- Dominici-Arosemena, A., and M. Wolff. 2005. Reef fish community structure in Bocas del Toro (Caribbean, Panamá): Gradients in habitat complexity and exposure. Caribbean Journal of Science 41(3):613–637.
- DuBar, J. R. 1974. Summary of the Neogene stratigraphy of southern Florida. Pp. 206–231 *in* R. Q. Oaks and J. R. DuBar, eds. Post-Miocene Stratigraphy Central and Southern Atlantic Coastal Plain. Utah State University Press, Logan, Utah.
- Dye, T. S., and K. R. Longenecker. 2004. Manual of Hawaiian Fish Remains Identification Based on the Skeletal Reference Collection of Alan C. Ziegler and Including Otoliths. Society for Hawaiian Archaeology Special Publication 1, Honolulu. 134 pp. Available for download at http://hawaiianarchaeology.org/publication/manual-of-hawaiian-fish-remains-identification-based-on-the-skeletal-reference-collection-of-alan-c-ziegler-and-including-otoliths-dye-and-longenecker-2004/.
- Emslie, S. D., W. D. Allmon, F. J. Rich, J. H. Wrenn, and S. D. de France. 1996. Integrated taphonomy of an avian death assemblage in marine sediments from the late Pliocene of Florida. Palaeogeography, Palaeoclimatology, Palaeoecology 124:107–136.
- Eschmeyer, W. N., R. Fricke, and R. van der Laan (eds.). Catalog of fishes: genera, species, references Retrieved from http://researcharchive.calacademy.org/research/Ichthyology/catalog/fishcatmain.asp on January 5, 2017.
- Food and Agriculture Organization of the United Nations Major Fishing Areas. Atlantic, western central (Major Fishing Area 31). Coordinating Working Party in Fishery Statistics (CWP) Data Collection. *In* Food and Agriculture Organization Fisheries and Aquaculture Department. Rome. Updated January 31, 2017. Retrieved from http://www.fao.org/geonetwork/srv/en/main.home?uuid=fao-fsa-map-31 on February 20, 2017.
- Fierstine, H., R. Huddleston, and G. Takeuchi. 2012. Catalog of the Neogene bony fishes of California, a systematic inventory of all pub-

- lished accounts. Occasional Papers of the California Academy of Sciences 159:1–206.
- Gašparič, R., R. Fraaije, B. van Bakel, J. Jagt, and P. Skupien. 2015. Mesozoic-Cenozoic crustaceans preserved within echinoid tests and bivalve shells. Bulletin of Geosciences 90(3):601–611.
- Gingerich, P. D., D. E. Russell, D. Sigogneau-Russell, J.-L. Hartenberger, S. M. I. Shah, M. Hassan, K. D. Rose, and R. H. Ardrey. 1979. Reconnaissance survey and vertebrate paleontology of some Paleocene and Eocene formations in Pakistan. Contributions from the Museum of Paleontology, University of Michigan 25:105–116.
- Glimore, G., and T. Fraser. 2015. *Apogon townsendi*. The International Union for Conservation of Nature (IUCN) Red List of Threatened Species. Retrieved from http://www.iucnredlist.org/details/185936/0 on January 11, 2017.
- Günther, A. 1859. Catalogue of the fishes in the British Museum. Catalogue of the acanthopterygian fishes in the collection of the British Museum: Gasterosteidae, Berycidae, Percidae, Aphredoderidae, Pristipomatidae, Mullidae, Sparidae 1:1–524.
- Halpern, B. S., and Floeter, S. R. 2008. Functional diversity responses to changing species richness in reef fish communities. Marine Ecology Progress Series 364:147–156.
- Harborne, A., H. Jelks, W. F. Smith-Vaniz, and L. Rocha. 2012. Abiotic and biotic controls of cryptobenthic fish assemblages across a Caribbean seascape. Coral Reefs 31:977–990.
- Harvey, J., T. Loughlin, M. Perez, and D. Oxman. 2000. Relationship between fish size and otolith length for 63 species of fishes from the eastern North Pacific Ocean. National Oceanic and Atmospheric Administration Technical Report National Marine Fisheries Service 150:1–36.
- Heilprin, A. 1886. Explorations on the west coast of Florida and in the Okeechobee wilderness. Transactions of the Wagner Free Institute of Science of Philadelphia 1:1–64.
- Hieb, K. 2000. Bay goby. Pp 139-141 in P. Olofson,

- ed. Bay lands ecosystem species and community profiles: life histories and environmental requirements of key plants, fish, and wildlife. Oakland, California.
- Huddleston, R., and G. Takeuchi. 2002. First Tertiary record of *Seriphus* (Perciformes: Sciaenidae) based on otoliths from the late Miocene of California. Journal of Vertebrate Paleontology 22(3):68a.
- Huddleston, R., and G. Takeuchi. 2006. A new late Miocene species of sciaenid fish, based primarily on an *in situ* otolith from California. Bulletin of the Southern California Academy of Sciences 105(1):30–42.
- Jawad, L., J. Al-Mamry, and H. Al-Busaidi. 2011. Relationship between fish length and otolith length and width in the lutjanid fish, *Lutjanus bengalensis* (Lutjanidae) collected from Muscat City coast on the Sea of Oman. Journal of Black Sea/Mediterranean Environment 17(2):116–126.
- Kemp, D. 1985. The Selsey Division (Bracklesham Group) at Lee-on-the-Solent, Gosport, (Hants). Tertiary Research 7(2):35–44.
- Kolbe, S. E., R. Lockwood, and G. Hunt. 2011. Does morphological variation buffer against extinction? A test using veneroid bivalves from the Plio-Pleistocene of Florida. Paleobiology 37(3):355-368
- Lacepède, B. G. E. 1801. Histoire naturelle des poissons 3:1–558.
- Lieske, E., and R. Myers. 1994. Collins Pocket Guide. Coral reef fishes. Indo-Pacific and Caribbean including the Red Sea. Harper Collins Publishers, 400 p.
- Mabuchi, K., T. H. Fraser, H. Song, Y. Azuma, and M. Nishida. 2014. Revision of the systematics of the cardinalfishes (Percomorpha: Apogonidae) based on molecular analyses and comparative reevaluation of morphological characters. Zootaxa 3846(2):151–203. doi:10.11646/zootaxa.3846.2.1.
- Mahadevan, S. 1961. The pearl fish *Carapus margaritiferae* (Rendahl), a new record for the Indian waters. Journal of Marine Biology Association of India 3(1–2):204–208.

- McEachran, J., and J. Fechhelm. 2005. Fishes of the Gulf of Mexico (Volume 2: Scorpaeniformes to Tetraodontiformes). University of Texas Press, Austin, Texas, 1004 p.
- Mendoza, R. P. 2006. Otoliths and their applications in fishery science. Ribarstvo 64(3):89–102.
- Nazar, A., G. Dharani, D. Rao, J. Santhanakumar, and N. Saravanane. 2011. A new record of pearl fish *Onuxodon margaritiferae* (Rendahl, 1921) from Andaman Islands. Indian Journal of Fisheries 58(2):141–143.
- Nelson, J. S., T. C. Grande, and M. Wilson. 2016. Fishes of the World (5th edition). John Wiley and Sons, Hoboken, New Jersey, 707 p.
- Nolf, D. 1985 Otolithi Piscium. Pp. 1–145 *in* H. Schultze, ed. Handbook of Paleoichthyology, 10, Gustav Fischer Verlag, Stuttgart, Germany and New York, United States.
- Nolf, D. 2013. The Diversity of Fish Otoliths, Past and Present. Operational Directorate "Earth and History of Life" of the Royal Belgian Institute of Natural Sciences, Brussels, Belgium, 581 p.
- Nolf, D., and S. Bajpai. 1992. Marine middle Eocene fish otoliths from India and Java. Bulletin de L'institut Royal des Sciences Naturelles de Belgique Sciences de la Terre 62:195–221.
- Parmentier, E., G. Castillo, M. Chardon, and P. Vandewalle. 2000. Phylogenetic analysis of the pearlfish tribe Carapini (Pisces: Carapidae). Acta Zoologica 81:293–306
- Parmentier, E., and K. Das. 2004. Commensal vs. parasitic relationship between Carapini fish and their hosts: some further insight through δ^{13} C and δ^{15} N measurements. Journal of Experimental Marine Biology and Ecology 310:47–58.
- Plate, L. 1908. *Apogonichthys strombi* n. sp., ein symbiotisch lebender Fisch von den Bahamas. Zoologischen Anzeiger 33(12):393–399.
- Randall, J. 1967. Food habits of reef fishes of the West Indies. Studies in Tropical Oceanography 5:665–847.
- Randall, J. 1996. Caribbean reef fishes. Third Edition. T.F.H. Publications, Inc. Ltd., Hong Kong, 368 p.

- Rivaton, J., and P. Bourret. 1999. Les otoliths des poissons de l'Indo-Pacifique. Documents Scientifiques et Techniques 112: 1–372.
- Robins, C., and G. Ray. 1986. A field guide to Atlantic coast fishes of North America. The Peterson Field Guide Series. Houghton Mifflin Company, Boston, U.S.A., 354 p.
- Rypel, A. 2008. An inexpensive image analysis system for fish otoliths. North American Journal of Fisheries Management 28(1):193–197.
- Schwarzhans, W., G. Carnevale, A. Bannikov, S. Japundžić, and K. Bradić. 2016. Otoliths *in situ* from Sarmatian (Middle Miocene) fishes of the Paratethys. Part I: *Atherina suchovi* Switchenska, 1973. Swiss Journal of Palaeontology. doi:10.1007/s13358-015-0111-0.
- Schwarzhans, W., and W. Weiler. 1971. Ein ungewöhnlicher Fund von Otolithen "in situ" aus dem mitteloligozänen Meeressand des Mainzer Beckens. Senckenbergiana lethaea 52(5-6):529–535.
- Scudder, S. J., E. H. Simons, and G. S. Morgan. 1995. Osteichthyes and Chondrichthyes from the Leisey Shell Pit local fauna, Hillsborough County, Florida. Bulletin of the Florida Museum of Natural History 37:251–272.
- Smith, J. L. B. 1955. The fishes of the family Carapidae in the West Indian Ocean. Annual Magazine of Natural History 12(3):401–416.
- Snyder, D., and G. Burgess. 2016. Marine Fishes of Florida. Johns Hopkins Press, Baltimore, Maryland, 392 p.
- Stinton, F. 1980. Fish otoliths from the English Eocene. Palaeontographical Society Monograph 4:191–258.
- Stringer, G. 1992. Late Pleistocene—Early Holocene teleostean otoliths from a Mississippi River mudlump. Journal of Vertebrate Paleontology 12(1):33–41.
- Stringer, G. 2016. Evidence and implications of marine invertebrate settlement on Eocene otoliths from the Moodys Branch Formation of Montgomery Landing (Louisiana, U.S.A.). Cainozoic Research 16(1):3–12.
- Takeuchi, G., and R. Huddleston. 2008. *Genyonemus whistleri* new species, a late Miocene sciaenid fish from California. Natural History

- Museum of Los Angeles County Sciences Series 41:279–291.
- Taverne, L., and D. Nolf. 2010. Priacanthidae (Teleostei, Perciformes) from the Lede Sands (Middle Eocene, Belgium): osteology and otoliths. Bulletin de L'Institut Royal des Sciences Naturelles de Belgique Sciences de la Terre 80:187–243.
- Trott, L. 1970. Contribution of the biology of carapid fishes (Paracanthopterygian: Gadiformes). University of California Publications in Zoology 89:1–41.
- Trott, L. 1981. A general review of the pearlfishes (Carapidae: Gadiformes) collected from Puerto Galera, Mindoro, Philippines. Copeia 1981:839–843.
- Watt, J., G. Pierce, and P. Boyle. 1997. Guide to the identification of North Sea fish using premaxillae and vertebrae. International Council for the Exploration of the Sea Report 220:1-231.
- Zullo, V., and D. Chivers. 1969. Pleistocene symbiosis: pinnotherid crabs in pelecypods from Cape Blanco, Oregon. The Veliger 12:72–74.